

Yiqi Luo · Robert B. Jackson · Christopher B. Field
Harold A. Mooney

Elevated CO₂ increases belowground respiration in California grasslands

Received: 8 November 1995 / Accepted: 5 March 1996

Abstract This study was designed to identify potential effects of elevated CO₂ on belowground respiration (the sum of root and heterotrophic respiration) in field and microcosm ecosystems and on the annual carbon budget. We made three sets of respiration measurements in two CO₂ treatments, i.e., (1) monthly in the sandstone grassland and in microcosms from November 1993 to June 1994; (2) at the annual peak of live biomass (March and April) in the serpentine and sandstone grasslands in 1993 and 1994; and (3) at peak biomass in the microcosms with monocultures of seven species in 1993. To help understand ecosystem carbon cycling, we also made supplementary measurements of belowground respiration monthly in sandstone and serpentine grasslands located within 500 m of the CO₂ experiment site. The seasonal average respiration rate in the sandstone grassland was 2.12 $\mu\text{mol m}^{-2} \text{s}^{-1}$ in elevated CO₂, which was 42% higher than the 1.49 $\mu\text{mol m}^{-2} \text{s}^{-1}$ measured in ambient CO₂ ($P = 0.007$). Studies of seven individual species in the microcosms indicated that respiration was positively correlated with plant biomass and increased, on average, by 70% with CO₂. Monthly measurements revealed a strong seasonality in belowground respiration, being low (0–0.5 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ in the two grasslands adjacent to the CO₂ site) in the summer dry season and high (2–4 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ in the sandstone grassland and 2–7 $\mu\text{mol CO}_2 \text{ m}^{-2} \text{ s}^{-1}$ in the microcosms) during the

growing season from the onset of fall rains in November to early spring in April and May. Estimated annual carbon effluxes from the soil were 323 and 440 g C m⁻² year⁻¹ for the sandstone grasslands in ambient and elevated CO₂. That CO₂-stimulated increase in annual soil carbon efflux is more than twice as big as the increase in aboveground net primary productivity (NPP_a) and approximately 60% of NPP_a in this grassland in the current CO₂ environment. The results of this study suggest that below-ground respiration can dissipate most of the increase in photosynthesis stimulated by elevated CO₂.

Key words Carbon cycle · Ecosystem · Global change · Respiration

Introduction

Belowground respiration is one of the primary pathways through which terrestrial ecosystems exchange carbon with the atmosphere. Studying belowground respiration as influenced by elevated CO₂ is essential for understanding carbon balance in terrestrial ecosystems in the future CO₂-enriched environment. In the past 15 years, experimental studies have indicated that photosynthetic carbon uptake is generally increased for plants grown in elevated CO₂ (Strain and Cure 1985; Luo et al. 1994). For example, photosynthesis averaged 44% higher for 39 tree species grown in elevated than in ambient CO₂ (Gunderson and Wullschlegel 1994). Additional carbon uptake in elevated CO₂ can stimulate carbon allocation to soil compartments through increased root biomass and exudation (Norby et al. 1987), accelerated root turnover rates (Rogers et al. 1994), and greater litterfall (N.R. Chiariello, unpublished data). Increased root biomass growth may respire more carbon, and increased availability of soil carbon may stimulate microbial activity in decomposing litter, all potentially leading to increased belowground respiration.

CO₂ effects on belowground respiration have been found to vary among ecosystems. Belowground respira-

CIWDPB Publication # 1271

Y. Luo (✉)¹ · R. B. Jackson² · H. A. Mooney
Department of Biological Sciences, Stanford University,
Stanford, CA 94305, USA

C. B. Field
Department of Plant Biology, Carnegie Institution of Washington,
Stanford, CA 94305, USA

Present addresses:

¹ Biological Sciences Center, Desert Research Institute,
P.O. Box 60220, Reno, NV 89506, USA,
fax: (702) 673-7485, e-mail: yluo@maxey.dri.edu

² Department of Botany, University of Texas at Austin,
Austin, TX 78713, USA

tion in the Alaskan tundra was not significantly affected by either long- or short-term CO₂ enrichment (Oberbauer et al. 1986). Belowground respiration at 525 ppm CO₂ was 50% higher than at 350 ppm in a ponderosa pine forest near Placerville, California, whereas 700 ppm CO₂ resulted in little change in belowground respiration for all three levels of nitrogen fertilization (Johnson et al. 1994; Vose et al. 1995). In a free-air CO₂ enrichment (FACE) experiment with cotton at the Maricopa Agricultural Center, Arizona, CO₂ enrichment at approximately 550 ppm significantly increased belowground respiration (Nakayama et al. 1994). Variation in CO₂ effects on belowground respiration may be related to other ecosystem attributes and processes. Long-term CO₂ enrichment in the Alaskan tundra, for example, did not stimulate ecosystem photosynthetic carbon fixation (Tissue and Oechel 1987; Grulke et al. 1990), whereas the FACE experiment with cotton led to changes in photosynthetic carbon uptake, soil carbon content (Wood et al. 1994), and root growth (Rogers et al. 1993; Prior et al. 1994).

Soil carbon dioxide is released to the atmosphere by respiration of living roots and microbial decomposition of litter added to soil from aboveground and belowground sources, including root exudates (Singh and Gupta 1977; Bowden et al. 1993). Annual belowground respiration rates correlate positively with aboveground litter production in forest ecosystems (Raich and Nadelhoffer 1989) and with mean productivity in different vegetation biomes on a global scale (Raich and Schlesinger 1992). Studies in specific ecosystems indicate that belowground respiration is also influenced by soil water content, temperature, and plant phenology (Singh and Gupta 1977). Seasonal patterns of belowground respiration, for example, are strongly associated with variations in soil water content in a tall prairie grassland (Grahammer et al. 1991) and with soil temperature and root activity in a Tennessee upland oak forest (Hanson et al. 1993).

This study is part of the Jasper Ridge CO₂ project designed to explore ecosystem-scale responses to elevated CO₂ (Field et al. 1996). The objective of this work was to identify the effect of increased CO₂ on belowground respiration in natural grasslands. We made three sets of measurements of belowground respiration: (1) seasonal time courses throughout one growing season in a sandstone grassland and in microcosms with mixed species from November 1993 to June 1994; (2) at peak biomass in both the serpentine and sandstone grassland in 1993 and 1994; and (3) at peak biomass in microcosms tubes with seven individual species in 1993. To help interpret and understand CO₂ effects on ecosystem carbon cycling, we made supplementary measurements to characterize seasonal patterns of belowground respiration, soil temperature, water content, and organic matter in the sandstone and serpentine grasslands adjacent to the CO₂ site. Based on the seasonal patterns and a simple model we developed, we estimate annual carbon fluxes and the ratio of annual belowground respiration to ecosystem productivity for the two CO₂ treatments and in the two natural grasslands.

Materials and methods

Site description

The CO₂ experiments, including microcosms and the two natural grasslands, are located at the Jasper Ridge Biological Preserve of Stanford University, California, (37° 24' N, 122° 14' W; elevation 150 m). The site has a mediterranean-type climate, wet in winter and dry in summer. During the last 20 years, annual precipitation varied from 200 mm (in 1975–1976) to 1200 mm (in 1982–1983), with an average of 595 mm from 1975 to 1990. The rainy season starts in fall, usually October or November, and lasts approximately until April (Chiariello 1989).

Soil properties of the two grasslands are listed in Table 1. The serpentine grassland at Jasper Ridge occupies serpentine-derived soils (Montara Series: loamy, serpentinitic, thermic, lithic haploxerolls) and sandstone grassland occupies sandstone-derived soil (Dibble Series, Millsholm variant: loamy, mixed, thermic, lithic xerochrepts; Kashiwagi 1985). Soil of the serpentine grassland has higher pH, cation exchange, soil nitrogen, and organic matter than the sandstone grassland soil but lower soil potassium (Table 1). The sandstone grassland is dominated by *Avena barbata* Link and *Bromus hordeaceus* L. in spring and *Hemizonia congesta* DC. ssp. *luzulifolia* (DC.) Bab. & H.M. Hall (Hickman 1993) in summer and early fall. Common species in the serpentine grassland include *Bromus hordeaceus*, *Calycadenia multiglandulosa* DC., *Lasthenia californica* Lindley, and *Plantago erecta* E. Morris (McNaughton 1968; Hobbs and Mooney 1985; Hickman 1993).

CO₂ experiments

The CO₂ experiments in the field used 30 plots of 0.33 m² soil area (0.65 m in diameter) randomly selected in each of the sandstone and serpentine grasslands for three treatments: ten replicates of no-chamber controls, open-top chambers with ambient CO₂, and open-top chambers with elevated CO₂ (ambient +350 ppm, a seasonal average of 723 ppm). Each cylindrical open-top chamber was 1 m tall and CO₂ fumigation started in January 1991. We focused on the two chamber treatments to examine CO₂ effects on belowground respiration.

For the microcosm experiments, 20 open-top chambers at either ambient or ambient+350 ppm CO₂ each contained an array of approximately 30 tubes which were 0.2 m in diameter and 1 m deep. The tubes were filled with an upper 0.15 m of shredded serpentine topsoil and 0.80 m of crushed rock from a serpentine quarry in 1992 and half of the tubes were filled with sandstone soil in 1993. We added one 38-mm watering in November of 1992 to ini-

Table 1 Soil Properties [pH, cation exchange (CE), Phosphorus (P), Potassium (K), nitrogen (N), soil organic matter (SOM), and texture (mean ± Se)] of the sandstone and serpentine grasslands. Kjeldahl nitrogen was analyzed as in Bremner and Mulvaney (1982), and P and K were extracted with 0.5 M sodium bicarbonate. SOM was measured by combusting dried soil samples at 500°C for 5 h. The data were averages from 0–3 cm surface soil for SOM and from 0–20 cm soil for the other attributes. Sample size was 3 for pH, CE, P, K, and N and 39 for SOM. *P* is a value of probability for the comparative *t* test from the sandstone and serpentine grasslands

Soil Properties	Sandstone	Serpentine	<i>P</i>
pH	5.5 ± 0.08	6.6 ± 0.03	0.002
CE (mmhos cm ⁻¹)	0.1 ± 0.00	0.7 ± 0.14	0.001
P (mg kg ⁻¹)	1.8 ± 0.09	1.8 ± 0.11	0.952
K (mg kg ⁻¹)	71 ± 2.2	44 ± 5.1	0.005
N (%)	0.10 ± 0.00	0.14 ± 0.01	0.004
SOM (%)	5.2 ± 0.1	7.5 ± 0.4	0.001
Texture	Loam	Clay loam	

tiate germination and did not add any supplemental water during the experiment. Monocultures of seven annual species common to Jasper Ridge grasslands were grown in the 1992–1993 season. The seven species included four C_3 grasses, *Avena barbata*, *Bromus hordeaceus*, *Lolium multiflorum* L., and *Vulpia microstachys* (Nutt.) Benth var. *pauciflora* (Beal) Lonard & Gould and three C_3 forbs, *Hemizonia congesta*, *Calycadenia multiglandulosa*, and *Plantago erecta* (Hickman 1993). In the 1993–1994 season, we grew two communities, a serpentine mixture [including *Vulpia microstachys*, *Lasthenia californica*, *Plantago erecta*, *Calycadenia multiglandulosa*, and *Lotus wrangelianus* Fischer & C. Meyer (Hickman 1993)] and a sandstone mixture (including *Avena barbata*, *Bromus hordeaceus*, *Hemizonia congesta*, and *Lotus wrangelianus*). The density for all treatments was 100–200 plants per tube, depending on species.

CO₂ fluxes measurements

Soil surface respiration was measured with a modified Li-Cor LI 6200 closed gas-exchange system (Li-Cor, Lincoln, Neb., USA) similar to that of Norman et al. (1992). The system provides belowground respiration values comparable to those measured by eddy correlation apparatus and potassium hydroxide traps (Norman et al. 1992; Dugas 1993). An open-ended cylinder was fitted with a Li-Cor sensor housing (Part No. 9960–035) with gas inlet and outlet ports. The belowground respiration chamber was 20.25 cm high and 4.10 cm in diameter, with a measurement surface area of 13.20 cm² and a chamber volume of 267.3 cm³. A leakage port in the upper part of the chamber maintained pressure equilibrium between the air inside and outside the chamber. The chamber was pressed slightly into the soil to ensure a good seal at the chamber-soil interface.

We avoided measurements within 5 days after rain because Norman et al. (1992) reported, and our preliminary data indicated, that soil degassing immediately after rain may result in overestimation of belowground respiration at the water-saturated soil surface. We did not make measurements under other atypical weather conditions (e.g., cloudy days), either, to avoid bias estimation of annual carbon flux from soil. We measured soil temperature at a depth of 3 cm below the soil surface with Type-T thermocouples connected to a data logger (Campbell 21 ×) and multiplexer. Soil samples in the surface 3-cm soil were taken for moisture measurement in the two grassland ecosystems. Soil water content was determined gravimetrically at 80°C.

Four to ten measurements were taken for each of the CO₂ treatments and in each of the two grassland ecosystems on each sampling date. For the supplementary measurements adjacent to the CO₂ site, individual measurement sites were randomly distributed throughout each system to incorporate natural spatial heterogeneity. All measurements were taken between 1100 and 1400 hours PST to minimize diurnal variability. The sampling time interval for these experiments to characterize time-courses of belowground respiration was approximately 1 month.

Estimation of the annual carbon flux from soil

Our estimates of annual total carbon efflux from the soil assume that our monthly measurements captured the broad seasonal patterns in below-ground respiration and that our midday measurements were approximate daily maxima. The latter were adjusted by soil temperature and a Q_{10} relationship to estimate daily efflux, which is used to estimate annual carbon efflux by multiplying the number of days (about 30 days) between measurement intervals.

Belowground respiration is generally predicted by both soil temperature and soil water content (Kim and Verma 1992). Due to little diurnal variation in soil water content, our estimates of daily carbon efflux were based on soil temperature. Belowground respiration at time h [$R(h)$] is estimated by

$$R(h) = \frac{R_{\max}}{Q_{10}^{[T_{\max} - T(h)]/10}}$$

where Q_{10} is a quotient indicating the increase in belowground respiration caused by a temperature increase of 10°C, R_{\max} is the daily maxima of belowground respiration, equaling our measured midday values, T_{\max} is the daily maximum temperature (°C) and $T(h)$ is the temperature at time h (°C), estimated by a general diurnal temperature oscillation equation (Monteith and Unsworth 1990). That is

$$T(h) = \frac{T_{\max} - T_{\min}}{2} \sin\left(2\pi \frac{h-6}{24}\right) + \frac{T_{\max} + T_{\min}}{2}$$

where T_{\min} is the daily minimum temperature (°C). Our measured values of midday soil temperature at the depth of 3 cm are used as T_{\max} and T_{\min} was the minimum soil temperature obtained from the weather station about 500 m away from the experimental sites.

Q_{10} values compiled by Raich and Schlesinger (1992) range from 1.3 to 3.3. We also performed a brief experiment to estimate a Q_{10} value in the sandstone grassland as a check on the model assumptions. Our measurements of belowground respiration, temperature, and water content started at 0900 hours on 6 March and ended at 0100 hours on 7 March 1993, with values taken every 2 h. Both belowground respiration and temperature in the two ecosystems were highest at midday and lowest at night while soil water varied little (data not presented). Belowground respiration increased as soil temperature increased with a Q_{10} value of 1.5, indicating that belowground respiration increased approximately 50% when soil temperature increased by 10°C. The measured Q_{10} value suggests that the relationship of belowground respiration with temperature in these grasslands may be close to the lower end of Raich and Schlesinger's (1992) Q_{10} range. In order to account for possibly different Q_{10} relationships in other seasons, we bracket our estimated daily and annual carbon fluxes for a range of Q_{10} between 1.5 and 2.5. Effects of seasonal variation in soil water content and temperature on belowground respiration were assumed to be reflected in our monthly measurements. Note that this simple model does not account for temperature gradient in the soil profile. Sensitivity analysis with Q_{10} ranging from 1.5 to 2.5, however, may bound variation in the annual carbon flux caused by the temperature gradient.

Results

CO₂ effects in field plots

Elevated CO₂ increased belowground respiration in the sandstone grassland from December to April, the active growing season for these plants (1993–1994, Fig. 1, $P = 0.007$). Belowground respiration was similar for the two CO₂ levels at the beginning or the end of the growing season, when the soil was dry and plant and microbial activity were minimal. In comparison to that in ambient CO₂, belowground respiration in elevated CO₂ increased 35% in December, about 60% in January and February, and 130% in March, when biomass approached its seasonal peak. The seasonal average of belowground respiration was 2.12 $\mu\text{mol m}^{-2} \text{s}^{-1}$ in elevated CO₂, which is significantly higher than 1.49 $\mu\text{mol m}^{-2} \text{s}^{-1}$ in ambient CO₂ ($P = 0.007$ by repeated measures analysis).

Relative effects of elevated CO₂ on belowground respiration were higher in 1994 than in 1993 in both sandstone and serpentine grasslands. In the sandstone grassland, where peak plant biomass occurs from March to May, belowground respiration was 43% ($P = 0.193$) higher in elevated CO₂ than that in ambient CO₂ in 1993 (Fig. 2A), 134% higher in March but much less stimulated in April and May in 1994 (Fig. 1). In the serpentine

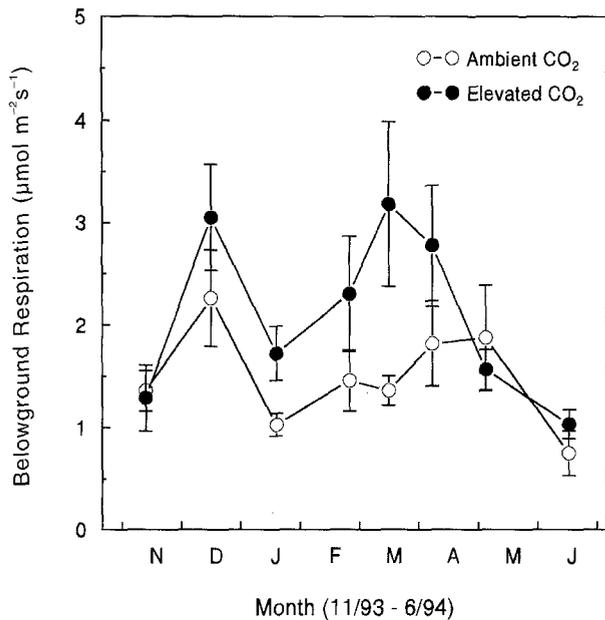


Fig. 1 Seasonal courses of belowground respiration in the sandstone grassland in ambient (*open circles*) and elevated CO₂ (*solid circles*; mean \pm SE, $n = 6-10$) from November 1993 to June 1994. Measurements were initiated in mid-November, a few days after the first major precipitation in fall, and continued though plant senescence in mid-June. Measurements were taken on bare spots underneath canopy in each field plot (10 plots per treatment)

grassland when peak biomass generally occurs in March and April, belowground respiration in elevated CO₂ was 36% higher ($P = 0.151$) in 1993 and 139% higher ($P = 0.062$) in 1994 than in ambient CO₂ (Fig. 2B). Differences between 1993 and 1994 were due primarily to a much drier growing season in 1994.

CO₂ effects in the microcosms

Time-course measurements in the microcosm experiment suggested little difference in belowground respiration between the two CO₂ levels before March 1994 (Fig. 3A, B). From March to May, belowground respiration in elevated CO₂ was 75% higher ($P = 0.046$) in the tubes filled with the sandstone soil and 60% higher ($P = 0.009$) with the serpentine soil than that in ambient CO₂. The seasonal average was not significantly different between the two CO₂ levels for the sandstone soil ($P = 0.788$), but approached significance for the serpentine ($P = 0.052$).

Studies of individual species in the microcosms indicated that below-ground respiration and its response to elevated CO₂ greatly varied with species (Fig. 4A). It was 1.5–2.5 $\mu\text{mol m}^{-2}$ ground area s^{-1} for *Plantago*, *Bromus*, *Hemizonia*, and *Calycadenia* and reached as high as 4–6 $\mu\text{mol m}^{-2}$ ground area s^{-1} for *Lolium*, *Avena*, and *Vulpia*. Elevated CO₂ had a significant effect on below-ground respiration in the tubes with *Plantago*, *Bromus*, *Hemizonia*, and *Lolium* but not for the other three species. The average of belowground respiration across species was 3.52 $\mu\text{mol m}^{-2}$ s^{-1} at elevated CO₂, nearly 70%

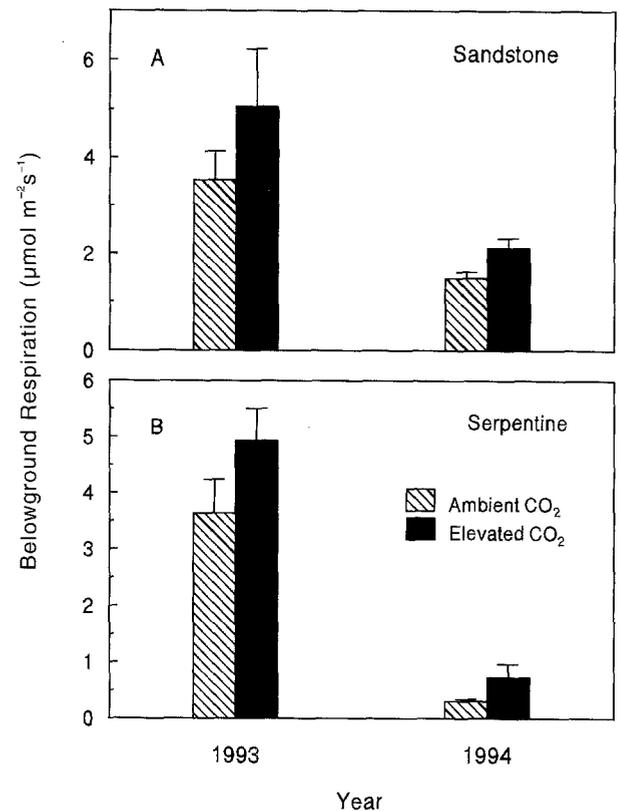


Fig. 2 Belowground respiration in the sandstone (A) and serpentine (B) grasslands in the ambient CO₂ (*hatched bars*) and elevated CO₂ (*solid bars*; mean \pm SE, $n = 4-10$) in 1993 and 1994. Measurements of belowground respiration were taken on the harvest rings immediately after cutting on 1 May 1993 in the sandstone grassland, on 5 April 1993 and 6 April 1994 in the serpentine grassland when plant biomass reached approximately the maximum. Data for the sandstone grassland in 1994 were the seasonal average from Fig. 1

greater than 2.10 $\mu\text{mol m}^{-2}$ s^{-1} at ambient CO₂ ($P = 0.001$). Belowground respiration in the tubes with monoculture of seven species was highly correlated with shoot biomass ($R^2 = 0.73$; data not shown) and total biomass ($R^2 = 0.65$) in the final harvest at the end of April and the beginning of May 1993 (Fig. 4B).

Annual carbon fluxes

In order to estimate annual carbon fluxes, we made monthly measurements of belowground respiration for 1 year in sandstone and serpentine grasslands located within 500 m of the CO₂ experiment site. Belowground respiration in the two natural grasslands increased from nearly zero in October to about 3 $\mu\text{mol m}^{-2}$ s^{-1} in November with the onset of fall rains (Fig. 5A, B). Respiration was maintained around 3 $\mu\text{mol m}^{-2}$ s^{-1} through the growing season from November to April. Summer belowground respiration was very close to zero (0–0.5 $\mu\text{mol m}^{-2}$ s^{-1} ; Fig. 5A, B), as most of the plants had senesced and little carbon was translocated below ground. Seasonal patterns of belowground respiration mirrored soil water availabil-

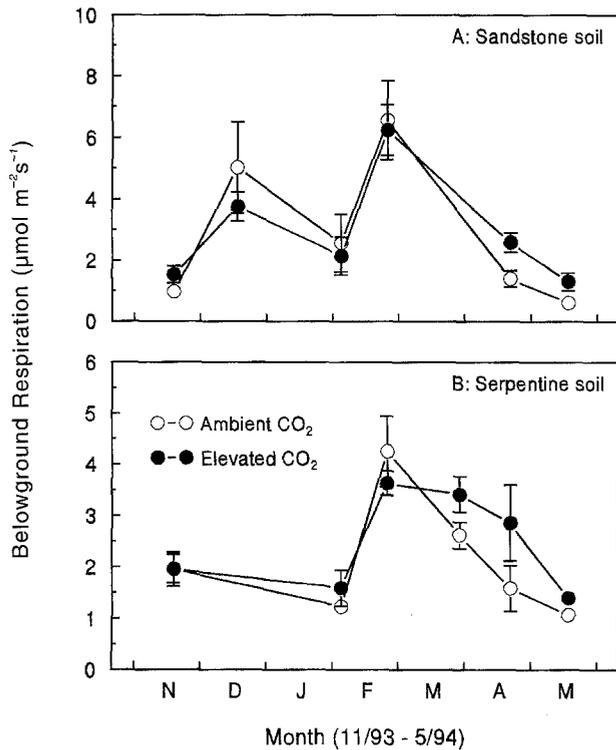


Fig. 3 Seasonal courses of belowground respiration in microcosms tubes with sandstone (A) and serpentine (B) soils in ambient (*open circles*) and elevated CO₂ (*solid circles*; mean \pm SE, $n = 4-6$) from November 1993 to May 1994. Measurements were initiated in mid-November and continued through peak biomass harvests in mid-May. Measurements were taken on bare spots between plants. Each microcosms tube was filled with well-mixed soils of either sandstone or serpentine at the beginning of growing season. Initial carbon contents in either sandstone or serpentine soils were presumably similar

ity quite closely (Fig. 5C, D). Belowground respiration was correlated positively with soil water content but negatively with soil temperature.

Using our model and the monthly measurements, we estimated that annual total carbon efflux from the soil of the sandstone grassland was 323 and 440 g C m⁻² year⁻¹, respectively, for the field plots with the ambient and elevated CO₂, differing by 36%, (Table 2, Q₁₀ = 2). Total carbon released from the sandstone and serpentine grasslands adjacent to the CO₂ site was 485 and 346 g C m⁻² year⁻¹. Annual carbon efflux from the CO₂ plots was less than that in the natural ecosystems partly because open-top chambers may alter microclimate (Drake and Peresta 1993; Lee and Barton 1993). Sensitivity analysis indicated that estimated annual carbon release decreases as Q₁₀ increases (this relationship occurs in our case because we estimate daily total carbon flux from a maximal respiration value at midday). When Q₁₀ varies from 1.5 to 2.5, estimated soil carbon was reduced by about 23%.

Based on the Raich and Nadelhoffer's (1989) approach, we also estimated the ratio of annual soil carbon efflux to aboveground net primary productivity which ranged from 2.07 to 7.69 for the two natural grasslands and with the two CO₂ treatments (Table 2). The ratio was

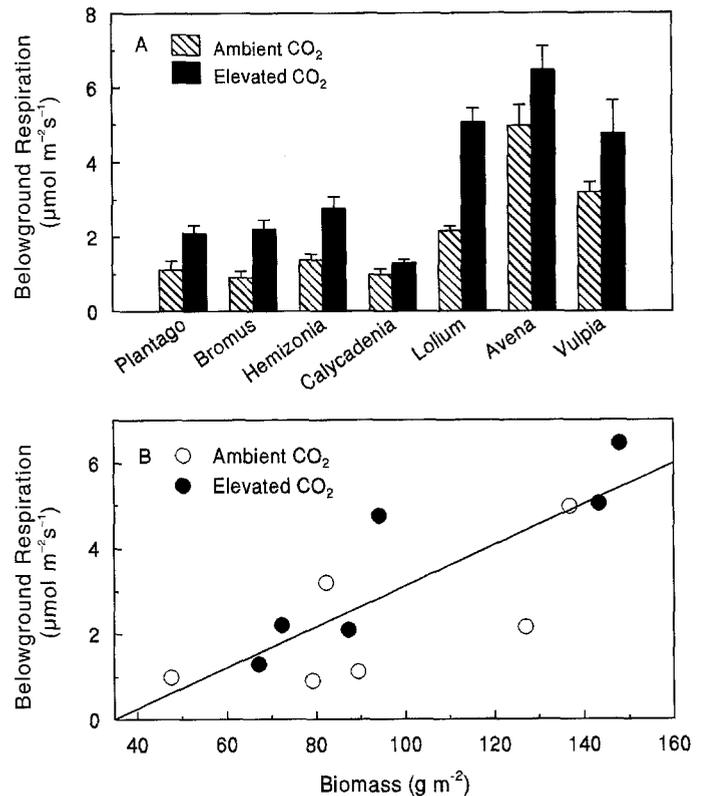


Fig. 4 A Belowground respiration in monoculture of seven species in microcosms tubes with ambient (*hatched bars*) and elevated CO₂ (*solid bars*; mean \pm SE, $n = 4-7$). Each tube was filled with well-mixed serpentine soils at the beginning of growing season. Measurements were taken in March and April 1993 when plant biomass reached the maximum. (B) The correlation between belowground respiration and total plant biomass (shoots plus roots) in the final harvest at the end of April and the beginning of May 1993 ($R^2 = 0.65$). No biomass data were available for *Calycadenia*

Table 2 Annual carbon release from soils to the atmosphere (R_{an} ; Q₁₀ = 2), aboveground net primary productivity (NPP_a), and the R_{an}/NPP_a ratio with the two CO₂ treatments in the sandstone grassland and in the sandstone and serpentine ecosystems outside the CO₂ site. R_{an} was estimated with our model and monthly measurements. Carbon efflux during the dry season from July to October in the plots with CO₂ treatments was assumed to be the same as in the natural ecosystem adjacent to the CO₂ site and was very close to zero. Values of NPP_a for the CO₂ treatments were estimated from data of live and dead biomass harvested in January and May 1994 (Field et al. 1996). Estimates of NPP_a for the sandstone and serpentine grassland ecosystems are from McNaughton (1968) and data of live and dead biomass on the no-chamber control plots in the CO₂ experiment (Chiariello et al., unpublished data). For all estimates, plant biomass is assumed to contain 45% carbon

Treatment	R_{an} g C m ⁻² year ⁻¹	NPP_a g C m ⁻² year ⁻¹	R_{an}/NPP_a
CO ₂ concentrations in plots on the sandstone grassland			
Ambient	323	104	3.11
Elevated	440	153	2.88
Ecosystems outside of the CO ₂ site			
Sandstone	485	135-234	2.07-3.59
Serpentine	346	45-90	3.84-7.69

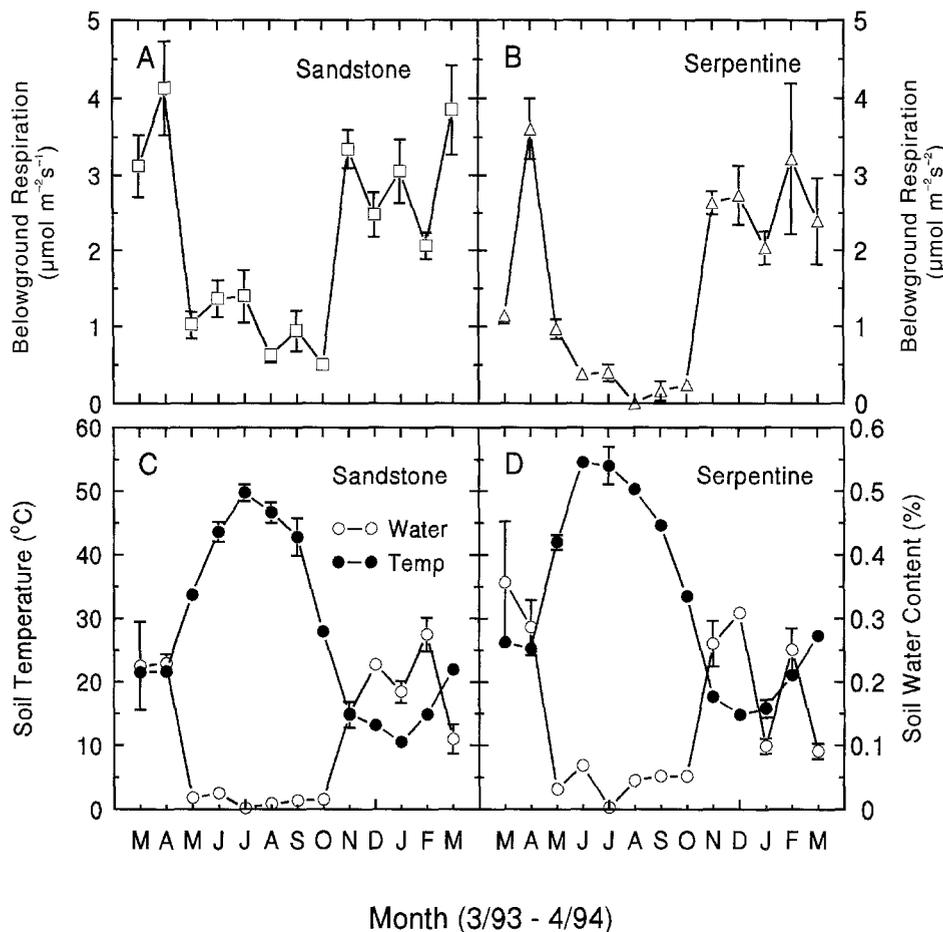


Fig. 5A, B Belowground respiration ($\mu\text{mol m}^{-2} \text{s}^{-1}$; mean \pm SE, $n = 5-7$), **C, D** soil temperature ($^{\circ}\text{C}$) at a depth of 3 cm (solid circles; mean \pm SE, $n = 3-6$), and gravitational water content (%) (open circles; mean \pm SE, $n = 3$) in the sandstone and serpentine grasslands located within 500 m of the CO_2 experiment site from March 1993 to March 1994. All measurements were made at noon between 1100 to 1400 hours PST once a month. Monthly summary of precipitation at the Jasper Ridge was 61.25, 22.10, 15.24, 0.00, 0.00, 0.00, 1.02, 15.74, 42.17, 96.27, 69.09, 163.58, and 7.62 mm, from March 1993 to March 1994, totaling 494.08 mm. The onset of fall rains occurred after our monthly respiration measurement in October. Belowground respiration (y) was positively correlated with soil water content (x) as $y = 1.14 + 8.87x$ ($R^2 = 0.54$; $P = 0.004$, where R^2 is determinant coefficient and P is a value of probability) for sandstone grassland and $y = 0.47 + 7.25x$ ($R^2 = 0.50$; $P = 0.007$) for serpentine grassland. Belowground respiration (y) is negatively correlated with soil temperature (x) as $y = 3.91 - 0.06x$ ($R^2 = 0.48$; $P = 0.009$) for sandstone grassland and $y = 3.86 - 0.07x$ ($R^2 = 0.66$; $P = 0.001$) for serpentine grassland

similar for the two CO_2 levels, indicating that elevated CO_2 altered aboveground productivity and belowground respiration proportionally. The ratio in the natural sandstone grassland ranged from 2.1 to 3.6, similar to values reported for other ecosystems. Values were much higher in the serpentine (3.8–7.7), possibly reflecting increased respiratory costs for growth and maintenance in stressful serpentine soils. Using different Q_{10} values between 1.5 and 2.5, the ratio varies only by about 20%.

Discussion

Our three sets of data all indicate that belowground respiration was higher in elevated than in ambient CO_2 . Elevated CO_2 increased the seasonal average of belowground respiration in the sandstone grassland by 42% in comparison to that in ambient CO_2 . Belowground respiration averaged across seven monoculture species in microcosm tubes was nearly 70% higher in elevated CO_2 than in ambient CO_2 . Although the seasonal average was not significantly different between the two CO_2 treatments in the microcosm tubes filled with either sandstone or serpentine soils, belowground respiration in the late growing season was significantly higher in elevated CO_2 than in ambient CO_2 . Elevated CO_2 increased belowground respiration at the peak biomass by 30–40% in 1993 and by about 130% in 1994 in both serpentine and sandstone grasslands. The yearly variation in CO_2 effects resulted from many factors, including a large difference in precipitation (905 mm in 1993 vs 433 mm in 1994) and consequent differences in plant growth and phenology (Jackson et al. 1995; Field et al. 1996).

Comparison of the seasonal course of CO_2 effects on belowground respiration in the sandstone grassland (Fig. 1) with those on the microcosm tubes (Fig. 3) suggests that elevated CO_2 may have altered soil carbon processes after the 2-year field CO_2 fumigation. The microcosm tubes were filled with well-mixed soils at the be-

ginning of the experiments, presumably having similar heterotrophic respiration for both CO₂ treatments. As a result, belowground respiration was not much different between the elevated and ambient CO₂ in the early growing season (Fig. 3A, B). Later in the growing season (March to May), plants grew larger and the contribution of root respiration to soil carbon fluxes may have been substantial as suggested by the correlation between biomass and respiration (Fig. 4B; Vose et al. 1995). Plant roots may respire some portion of increased carbon newly fixed in photosynthesis at elevated CO₂ (Bowden et al. 1993; Cheng et al. 1994), resulting in higher belowground respiration than at ambient CO₂ in the late growing season.

For plots with open-top chambers in the sandstone grassland, root respiration during the early growing season was unlikely to contribute much to belowground respiration because the plants were very small. Increased belowground respiration at elevated CO₂ may be attributable to litter feedback effects. Aboveground litter was increased by 44% in January 1993 and 30% in May 1994 in elevated CO₂ compared to ambient CO₂ (Field et al. 1996). Root biomass growth was increased by 26% (Hungate et al. 1996), leading to increased belowground litter in elevated CO₂. In addition, the lignin content of roots was reduced in elevated compared to ambient CO₂ (Chu et al. 1996), suggesting higher decomposability of soil organic matter. Increased carbon availability in soil and reduced lignin content may stimulate microbial population and activities. Indeed, soil microbial biomass increased by 55% in April 1992, 15% in January and 40% in May 1994 in elevated CO₂ (Hungate et al. 1996). Overall, elevated CO₂ in the sandstone grassland may have increased litterfall with less lignin which stimulated microbial population and heterotrophic respiration, resulting in higher belowground respiration in elevated than in ambient CO₂, even in the early growing season.

In the late growing season in the sandstone grassland, belowground respired carbon came partly from litter and partly from current photosynthesis (Bowden et al. 1993; Cheng et al. 1994). Photosynthetic carbon uptake of *Avena barbata*, the dominant species, was stimulated by up to 70% in comparison to that in ambient CO₂ in 1992–93 and 1993–94 growth season (Jackson et al. 1994, 1995). Modeling studies indicate that such an increase in leaf photosynthetic carbon uptake in elevated CO₂ may lead to a 30–110% increase in seasonal carbon influx into the system, depending on physiological adjustments in leaf death, allocation, and nonstructural carbon storage (Luo et al. 1996). In addition, increased soil water availability in elevated CO₂ (Field et al. 1995) may possibly amplify CO₂ effects on belowground respiration in the late growing season through enhanced plant growth (Field et al. 1996) and microbial decomposition (Hungate et al. 1996).

Annual soil carbon efflux in the sandstone grassland was estimated to increase by about 120 g m⁻² year⁻¹ in elevated CO₂ (Table 2). This CO₂-stimulated increase in soil carbon efflux is more than twice as large as the in-

crease in aboveground net primary productivity (NPP_a) and approximately 60% of NPP_a in this grassland in the current CO₂ environment. The increase in respiration indicates that a considerable amount of carbon was translocated below ground for root respiration, root turnover rate, and root exudation. Based on the general relationship between belowground respiration and NPP_a by Raich and Schlesinger (1989), Hungate et al. (1996) estimated that a combination of root respiration, turnover rate, and exudation was increased by 56% in the sandstone grassland in elevated CO₂ in comparison to that in ambient CO₂. In addition, that increase in belowground respiration may signify changes in carbon residence time in elevated CO₂. Total soil carbon content is estimated to be 8000 g C m⁻² in the sandstone grassland (Hungate et al. 1996). Carbon residence time is estimated to be nearly 25 years in ambient CO₂ with annual carbon efflux being 323 g m⁻² year⁻¹. Elevated CO₂ increased carbon fluxes to 440 g m⁻² year⁻¹ but did not significantly change soil carbon content (Hungate et al. 1996). Carbon residence time is consequently decreased to about 18 years in elevated CO₂, potentially reducing carbon sequestration capacity in the grassland ecosystem.

Overall, elevated CO₂ increased belowground respiration in comparison to that in ambient CO₂, consistently across different CO₂ experiments. Belowground respiration and its response to elevated CO₂ varied with plant biomass in the microcosms, exhibited a strong seasonality and were subjected to year-to-year variation. Comparative analysis on these sets of data from the microcosms and field CO₂ plots may suggest litter feedback effects on belowground respiration in the sandstone grassland after the 2-year CO₂ fumigation. Estimated annual carbon effluxes from the soil were increased by nearly 120 g C m⁻² year⁻¹, implying that belowground respiration was sufficiently large to dissipate most of the increase in photosynthetic carbon fixation stimulated by elevated CO₂.

Acknowledgements We thank F.S. Chapin and B.A. Hungate for use of the respiration chamber. The Jasper Ridge CO₂ Experiment is supported by grants from the U.S. National Science Foundation to the Carnegie Institution of Washington, Stanford University, and the University of California at Berkeley. R.B.J. was supported by a DOE Distinguished Postdoctoral Fellowship for Global Change. We thank two anonymous reviewers, B.A. Hungate, E.A. Hollend, D.W. Johnson, J. Canadell, and A. Austin for helpful comments on the manuscript and N. Chiariello for providing the weather data.

References

- Bowden RD, Nadelhoffer KJ, Boone RD, Melillo JM, Garrison JB (1993) Contributions of aboveground litter, belowground litter, and root respiration to total soil respiration in a temperate mixed hardwood forest. *Can J For Res* 23: 1402–1407
- Bremner JM, Mulvaney CS (1982) Nitrogen-total. In: Page AL (ed) *Methods of soil analysis. II*. American Society of Agronomy, Madison, Wis, pp 595–624
- Cheng W, Coleman DC, Carroll CR, Hoffman CA (1994) Investigating short-term carbon flows in the rhizospheres of different plant species, using isotopic trapping. *Agron J* 86: 782–788

- Chiariello NR (1989) Phenology of California grasslands. In: Huehneke LF, Mooney HA (eds) Grassland structure and function: California annual grassland. Kluwer, Dordrecht, pp 47–58
- Chu CC, Field CB, Mooney HA (1996) Effects of CO₂ and nutrient amendment on tissue quality of two California annuals. *Oecologia* (in press)
- Drake BG, Peresta GJ (1993) Open-top chambers for studies of the long-term effects of elevated atmospheric CO₂ and carbon balance on wetland and forest ecosystem processes. In: Schulze ED, Mooney HA (eds) Design and execution of experiments on CO₂ enrichment. Commission of the European Communities, Luxembourg, pp 273–289
- Dugas WA (1993) Micrometeorological and chamber measurements CO₂ flux from bare soil. *Agric For Meteorol* 67: 115–128
- Field CB, Jackson RB, Mooney HA (1995) Stomatal responses to increased CO₂: implications from plant to global scale. *Plant Cell Environ* 1214–1225
- Field CB, Chapin FS III, Chiariello NR, Holland EA, Mooney HA (1996) The Jasper Ridge CO₂ experiment: design and motivation. In: Koch GW, Mooney HA (eds) Carbon dioxide and terrestrial ecosystems. Academic Press, San Diego, CA, pp. 121–145
- Grahammer K, Jawson MD, Skopp J (1991) Day and night soil respiration from a grassland. *Soil Biol Biochem* 23: 77–81
- Grukke NE, Riechers GH, Oechel WC, Hjelm U, Jager C (1990) Carbon balance in tussock tundra under ambient and elevated atmospheric CO₂. *Oecologia* 83: 485–494
- Gunderson CA, Wullschlegel SD (1994) Photosynthetic acclimation in trees to rising atmospheric CO₂: a broader perspective. *Photosynth Res* 39: 369–388
- Hanson PJ, Wullschlegel SD, Bohlman SA, Todd DE (1993) Seasonal and topographic patterns of forest floor CO₂ efflux from an upland oak forest. *Tree Physiol* 13: 1–15
- Hickman JC (1993) *The Jepson Manual: higher plants of California*. University of California Press, Berkeley, Calif
- Hobbs RJ, Mooney HA (1985) Community and population dynamics of serotinous grassland annuals in relation to gopher disturbance. *Oecologia* 67: 342–351
- Hungate BA, Jackson RB, Field CB, Chapin FS (1996) Field CO₂ enrichment experiments lack statistical power to detect changes in soil carbon cycling. *Plant Soil* (in press)
- Jackson RB, Sala OE, Field CB, Mooney HA (1994) CO₂ alters water use, carbon gain, and yield for a dominant species in a natural grassland. *Oecologia* 98: 257–262
- Jackson RB, Luo Y, Cardon ZG, Sala OE, Field CB, Mooney HA (1995) Photosynthesis, growth, and density for the dominant species in a CO₂-enriched grassland. *J Biogeogr* 22: 221–225
- Johnson DW, Geisinger DR, Walker RF, Newman J, Vose JR, Elliot KJ, Ball JT (1994) Soil pCO₂, soil respiration, and root activity in CO₂-fumigated and nitrogen-fertilized ponderosa pine. *Plant Soil* 165: 129–138
- Kashiwagi J (1985) Soil map of the Jasper Ridge Biological Preserve. Soil Conservation Service Map. Jasper Ridge Biological Preserve Publication, Stanford, Calif
- Kim J, Verma SB (1992) Soil surface CO₂ flux in a Minnesota peatland. *Biogeochemistry* 18: 37–51
- Lee HSJ, Barton CVM (1993) Comparative studies on elevated CO₂ using open-top chambers, tree chambers and branch bags. In: Schulze ED, Mooney HA (eds) Design and execution of experiments on CO₂ enrichment. Commission of the European Communities, Luxembourg, pp 239–259
- Luo Y, Field CB, Mooney HA (1994) Predicting responses of photosynthesis and root fraction to elevated CO₂: interactions among carbon, nitrogen, and growth. *Plant Cell Environ* 17: 1195–1204
- Luo Y, Chen JL, Reynolds JF, Field CB, Mooney HA (1996) Elevated CO₂ altered plant carbon balance in California grasslands: modeling studies with a generic plant simulator (GePSi). *Ecol Model* (in press)
- McNaughton SJ (1968) Structure and function of California grasslands. *Ecology* 49: 962–972
- Monteith JL, Unsworth MH (1990) Principles of environmental physics, 2nd edn. Arnold, London
- Nakayama FS, Huluka G, Kimball BA, Lewin KF, Nagy J, Hendrey GR (1994) Soil carbon dioxide fluxes in natural and CO₂ enriched systems. *Agric For Meteorol* 70: 131–140
- Norby RJ, O'Neill EG, Hood WG, Luxmoore RJ (1987) Carbon allocation, root exudation and mycorrhizal colonization of *Pinus echinata* seedlings grown under CO₂ enrichment. *Tree Physiol* 3: 203–210
- Norman JM, Garcia R, Verma SB (1992) Soil surface CO₂ fluxes and the carbon budget of a grassland. *J Geophys Res* 97: 18845–18853
- Oberbauer SF, Oechel WC, Riechers GH (1986) Soil respiration of Alaskan tundra at elevated atmospheric carbon dioxide concentrations. *Plant Soil* 96: 145–148
- Prior SA, Rogers HH, Runion GB, Mauney JR (1994) Effects of free-air CO₂ enrichment on cotton root growth. *Agric For Meteorol* 70: 69–86
- Raich JW, Nadelhoffer KJ (1989) Belowground carbon allocation in forest ecosystems: global trends. *Ecology* 70: 1346–1354
- Raich JW, Schlesinger WH (1992) The global carbon dioxide flux in soil respiration and its relationship to vegetation and climate. *Tellus* 44B: 81–99
- Rogers HH, Prior SA, O'Neill EG (1993) Cotton root and rhizosphere responses to free-air CO₂ enrichment. In: Hendrey GR (ed) FACE: free-air CO₂ enrichment for plant research in the field. CRC Press, Boca Raton, Fla, pp 251–263
- Rogers HH, Runion GB, Krupa SV (1994) Plant responses to atmospheric CO₂ enrichment with emphasis on roots and the rhizosphere. *Environ Pollut* 83: 155–189
- Singh JS, Gupta SR (1977) Plant decomposition and soil respiration in terrestrial ecosystems. *Bot Rev* 43: 449–528
- Strain BR, Cure JD (1985) Direct effects of increasing carbon dioxide on vegetation. United States Department of Energy, DOE/ER-0238, Washington, DC
- Tissue DT, Oechel WC (1987) Response of *Eriophorum vaginatum* to elevated CO₂ and temperature in the Alaskan tussock tundra. *Ecology* 68: 401–410
- Vose JM, Elliot KJ, Johnson DW, Walker RF, Johnson MG, Tingey DT (1995) Effects of elevated CO₂ and N fertilization on soil respiration from ponderosa pine (*Pinus ponderosa*) in open-top chambers. *Can J For Res* 25: 1243–1251
- Wood CW, Torbert HA, Rogers HH, Runion GB, Prior SA (1994) Free-air CO₂ enrichment effects on soil carbon and nitrogen. *Agric For Meteorol* 70: 103–116